Novel Method of Acanthamoeba Movement Quantification to Determine Efficacy of Contact Lens Care Solutions

Introduction

Acanthamoeba keratitis is a serious ocular infection which is extremely challenging to treat.¹

May lead to permanent corneal damage and blindness.

Infection occurs when it comes in contact with contact lenses, usually via water, and is transported to the eye. Habits of Acanthamoeba

during both contamination and disinfection have remained elusive.

By quantifying Acanthamoeba motility, we may be able to better understand disinfection efficacy of contact lens products.

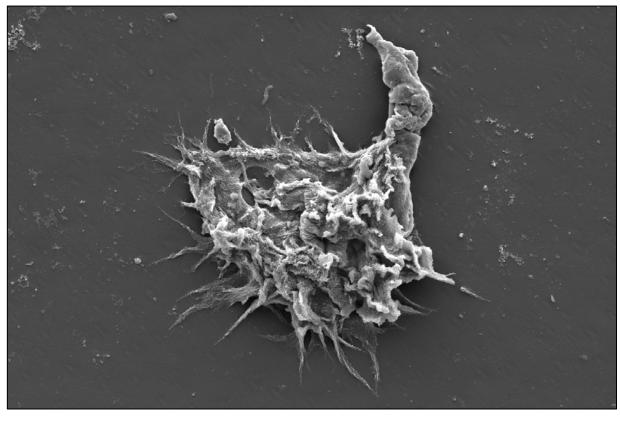


Figure 1. Acanthamoeba trophozoite, Alcon Research, LLC 202

Methods

Two Acanthamoeba strains were used (A. polyphaga, American Tissue Culture Collection (ATCC) 30461, and A. *castellanii*, ATCC 50370).

Acanthamoeba were scaled up 24 hours prior to testing in axenic culture media to create homogenous trophozoite populations.^{2, 3}

Acanthamoeba were seeded in a sterile aluminum flow cell at 7.5 x 10³ to 3x10⁴ CFU/mL and allowed to adhere to the surface for 30 minutes.

The flow cell solution was then changed to one of the following:

Control solutions:

- ¹⁄₄ Ringer's solution, on glass
- ¹⁄₄ Ringer's solution with heat-killed *E. coli*, on glass
- Axenic culture media (AC6), on glass
- Non-nutrient amoeba saline (NNA), on agar

Contact Lens Care Solutions (CLCs):

- <u>PHMB</u> (polyhexamethylene biguanide)
- <u>PAPB</u> (polyaminopropyl
- biguanide)/<u>Polyquad</u> (polyquaternium) <u>Polyquad/Aldox</u> (myristamidopropyl dimethylamine)
- <u>Polyquad/Alexidine</u>

Images taken once every 24 seconds during recording to create a for up to 12 hours at 4x magnification.

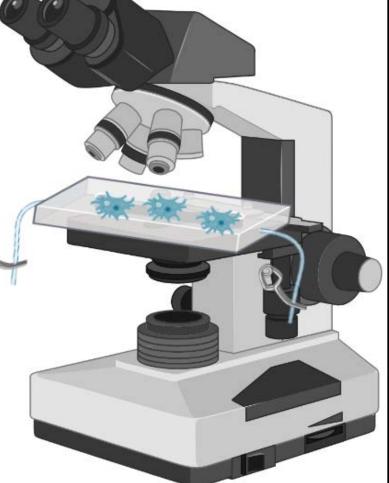
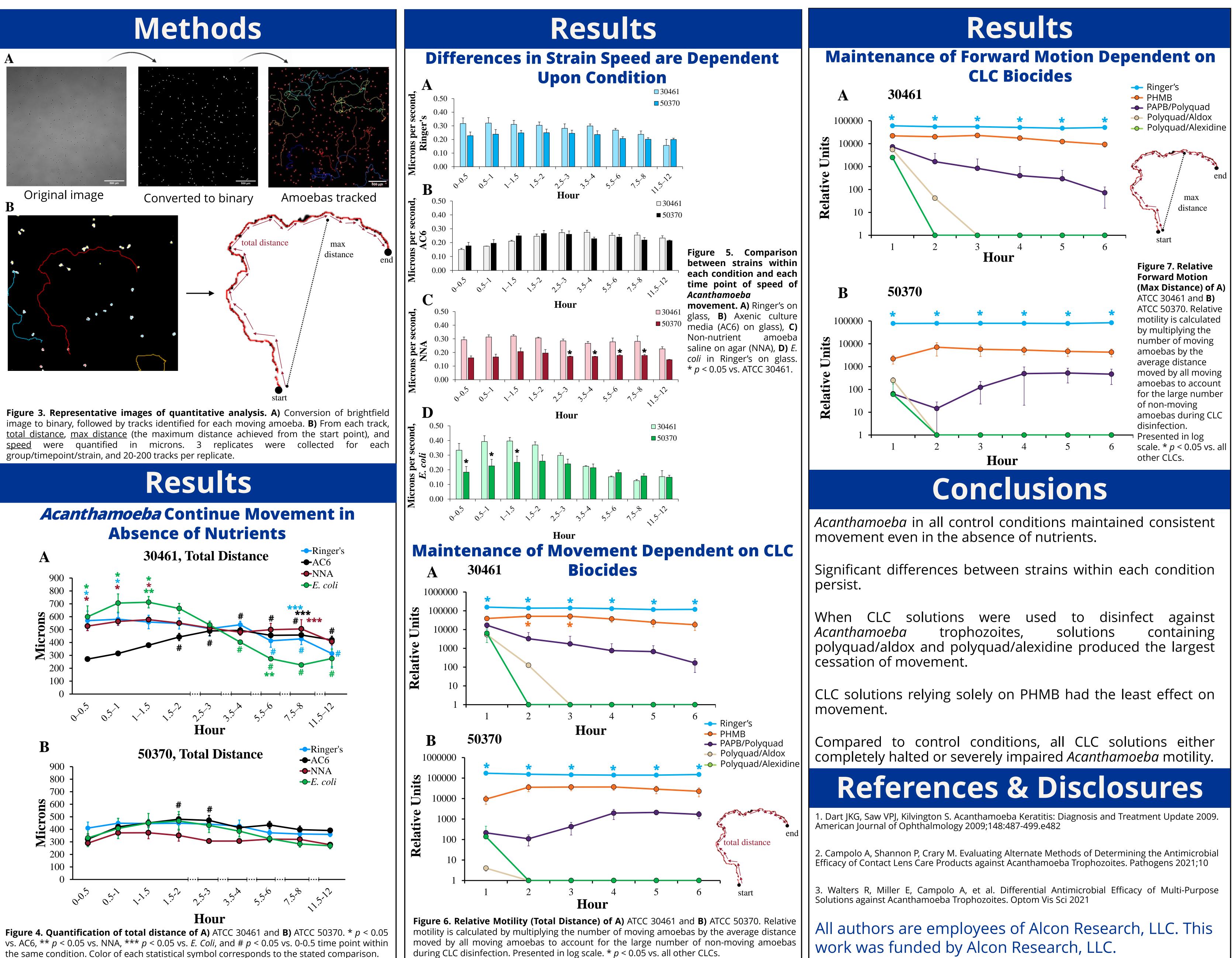


Figure 2. Depiction of sterile aluminum flow cell with Acanthamoeba visible through the glass coverslip. Both ends are clamped closed environment with no flow or shear stress.



Allison Campolo, Rhonda Walters, Brian Patterson, Monica Crary*



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* Corresponding author: +1 (817) 5518551, monica.crary@alcon.com